

Some problems in microscopy encountered in clinics for sexually transmitted disease

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SUMMARY The actual field diameter of 24 microscopes used in clinics for sexually transmitted diseases has been measured by means of a stage micrometer. The variation in findings is given, and the difficulties in producing accurate measurements are discussed.

Introduction

It is an accepted truth that accurate measurements of selected parameters are essential to the scientific study of problems. For this to be achieved it is necessary for the units of measurement to be constant in different laboratories and for different observers. In the days when the cubit was the accepted unit of length and was defined as the length of the king's forearm, difficulties could arise when a new king was crowned.

A commonly accepted method of assessing the severity of urethritis is to quote the number of leucocytes found in a 'high power field'. Unfortunately, this unit suffers from some of the disadvantages of the cubit.

The edge of the field of the microscope is an image of the stop within the eyepiece. The actual diameter of the field, expressed in micrometers, will depend on the total magnification which is the product of the eyepiece and objective magnifications, and in some instruments this must be increased by the magnification factor of the binocular head. A further complication is that eyepiece design may affect the field size; for example, a Huyghens $\times 8$ eyepiece may have a smaller field than a wide field $\times 10$ eyepiece.

Methods

By courtesy of the staff it has been possible to measure the actual field diameter of the microscopes used in five major clinics in central London and in two in the home counties. Measurements were carried out with a stage micrometer.

Results and comment

A total of 24 instruments was studied. The results are shown in the Table. The extreme variation of field diameter was from 112 μm to 190 μm , so that if the same specimen was studied a count of 20 cells in a field of 112 μm would be given as 57 cells per field of 190 μm .

For some of the instruments two values for the field size are given. In these microscopes the interocular distance of the binocular head is regulated by moving the eyepiece tubes along a horizontal slideway; this has the effect of varying the tube-length and, consequently, the magnification. This error is not too serious since the value of 20 cells per minimum field would only be reported as 24 cells per maximum field. In those instruments which have a single value the interocular distance is adjusted by pivoting the eyepiece tubes in the manner used for field-glasses; this mechanism maintains a constant tubelength and the magnification is unaltered.

A further complication is that in several of the instruments the outer zone of the field had very poor definition, which could not be improved by careful focusing, and was therefore due to lack of correction of spherical aberration in the outer zone of the objective.

It would seem that if cell counts are to serve any useful purpose the field size must be standardised. The simplest method would be to use a square field stop or graticule in the eyepiece, as in the Ehrlich eyepiece. This would have to be calibrated against a stage micrometer, a task which could be done by the service mechanics during routine maintenance. Hopefully the cost might not be prohibitive, and the venereologist's field would then have a more constant basis than the royal cubit.

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Table Detailed findings in 24 microscopes studied*

		Diameter of field (μm)		Area of field (μm²)			
		Single value	Double value		Single value	Double value	
Microscope no.	Eyepiece		Min.	Max.		Min.	Max.
1	Huyghens × 6	170			22 698		
2	Wide field × 6	172			23 235		
3	Periplan × 6		159	168		19 856	22 167
4	Huyghens × 6		172	190		23 235	28 353
5	Huyghens × 6		140	158		15 399	19 609
6	Huyghens × 8	125			12 272		
7	Huyghens × 8		173	185		23 506	26 880
8	Compensating × 8		155	165		18 869	21 383
9	Huyghens × 8	120			11 310		
10	Huyghens × 8		180	190		25 447	28 353
11	Huyghens × 8		175	180		24 053	25 447
12	Huyghens × 8		164	170		21 124	22 698
13	Huyghens × 8		115	120		10 387	11 310
14	Huyghens × 8		112	119		9852	11 122
15	Compensating × 8	138			14 957		
16	Wide field × 10	156			19 113		
17	Wide field × 10	161			20 358		
18	Periplan × 10		180	190		25 447	28 353
19	Periplan × 10		173	183		23 506	26 302
20	Compensating × 10	153			18 358		
21	Huyghens × 10		170	172		22 698	23 235
22	Compensating × 10		150	165		17 671	21 383
23	Wide field × 10		168	178		22 167	24 885
24	Compensating × 12.5	170			22 698		

*All the instruments were equipped with oil immersion objectives of simple achromatic type, except Nos. 20 and 24, which had flat field achromats. In all cases these were $\times 100$ magnification.

Further difficulties arise when the texture of the smears is considered. Such preparations are rarely uniform in cellular distribution. For this reason a quick survey of the whole smear should be made under a low power before any count is attempted. Since some microscopes are only equipped with an

oil immersion objective, this cannot always be done. Naturally, when the pressure of work is great there is a temptation for the technician to use the high power at once; if the smear is very scanty this may account for a few of the reports of 'nil on slide after staining'.